



ToyoScreen®

INTRODUCTION

Large scale commercial purifications begin at a small scale in methods development. In some cases the developers wish to use bulk resin to pack their initial small columns. By doing so, they experience how the resin handles physically and they can use this knowledge later during scale up as the column volumes become larger. In other cases the speed and convenience of having a pre-packed column for resin evaluation is needed. Tosoh Bioscience is pleased to offer pre-packed ToyoScreen Process Development Columns containing our popular TOYOPEARL® resins for evaluation. The ToyoScreen Series consist of small screening columns packed with TOYOPEARL, a packing material for semi-preparative and preparative liquid chromatography. These columns are suitable for evaluating different TOYOPEARL resins or for developing the purification conditions of biological target molecules such as proteins or nucleic acids. The Toyo-Screen Series is available in two column volumes (1 mL and 5 mL formats).

HIGHLIGHTS

- Packed with TOYOPEARL hydrophobic interaction, ion exchange, mixed mode or affinity chemistries.
- Low cost, efficient alternative to self packing.
- Easy connections with ÄKTA®, FPLC and HPLC.
- Offered in mixed or single chemistry packages of 5 or 6

SCREENING

TOYOPEARL, for example, is available in four different particle sizes and three different pore sizes. So optimal selection of a particular resin could involve screening of several resins.

HIC - HYDROPHOBIC INTERACTION

Hydrophobic Interaction Chromatography (HIC) sorts biomolecules by degree of their surface hydrophobicity. Samples are adsorbed to the resin at relatively high salt concentrations and eluted with a decreasing salt gradient. The mild conditions used in HIC separation typically maintain protein structure and biologic activity. Separation can either be optimized by varying the mobile phase or by using different HIC packings. TOYOPEARL HIC media are available in six different chemistries ranging in hydrophobicity from Ether-650 (low) to Hexyl-650 (high), see Figure 1. Depending on the target feedstock and impurity profile, the determination of the best selectivity is an empirical process. Figure 2 shows the selectivity differences of the ToyoScreen HIC chemistries on the separation of protein standards and antibodies from albumin in mouse ascites fluid.

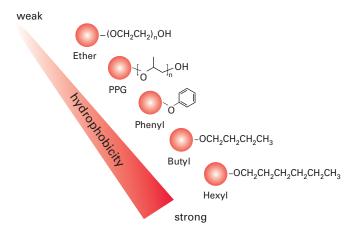
TOYOSCREEN SERIES PACKED WITH TOYOPEARL

TOYOPEARL	Particle size (µm)	IEX (eg/L-gel)	SBC (g/L-gel)
IEC type			
DEAE-650M	40 - 90	0.08 - 0.12	25 - 35 ¹⁾
GigaCap Q-650M	50 - 100	0.10 - 0.20	≥ 162 ¹⁾
SuperQ-650M	40 - 90	0.20 - 0.30	105 - 155 ¹⁾
Q-600C AR	50 - 150	0.15 - 0.20	> 1201)
QAE-550C	50 - 150	0.28 - 0.38	60 - 80 ¹⁾
GigaCap CM-650M	50 - 150	0.17 - 0.28	$\geq 110^{6)}$
NH2-750F	30 - 60	0.07 - 0.13	$\geq 70^{6)}$
GigaCap S-650M	50 - 100	0.10 - 0.20	136 - 176 ⁶⁾
SP-650M	40 - 90	0.13 - 0.17	40 - 602)
SP-550C	50 - 150	0.14 - 0.18	80 - 1202)
HIC type			
Ether-650M	40 - 90	-	10 - 30 ²⁾
Phenyl-650M	40 - 90	-	30 - 502)
Phenyl-600M	40 - 90	-	45 - 652)
Butyl-650M	40 - 90	-	30 - 502)
Butyl-600M		-	40 - 602)
Hexyl-650C	50 - 150	-	30 - 502)
PPG-600M	40 - 90	-	20 - 353)
SuperButyl-550C	50 - 150	-	52 - 70 ²⁾
MX-type			
MX-Trp-650M	50 - 100	-	> 756)
AFC type			
AF-rProtein A HC-650F	30 - 60	-	≥ 6 8 ⁶⁾
AF-rProtein A-650F	30 - 60	-	> 456)
AF-Chelate-650M	40 - 90	0.025 - 0.045	-
AF-Blue HC-650M	40 - 90	-	≥ 18 ⁴⁾
AF-RED-650M	40 - 90	-	2.5 - 4.54)

Measured with $^{1)}$ Bovine serum albumin, $^{2)}$ Lysozyme, $^{3)}$ $\gamma\text{-Globulin,}$ $^{4)}$ Human serum albumin, $^{5)}$ Antithrombin-III (Tosoh original method.), $^{6)}$ IgG

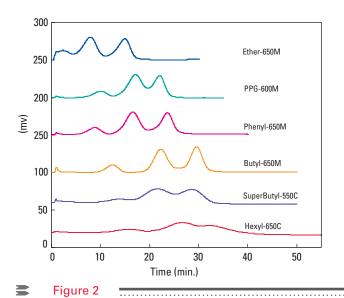
Table 1

HIC LIGAND CANDIDATES

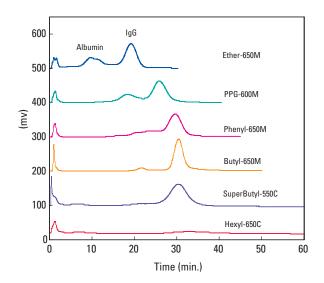




SCREENING OF TOYOPEARL HIC RESINS - STANDARD PROTEINS



SCREENING OF TOYOPEARL HIC RESINS - MOUSE ASCITES FLUID



Column: ToyoScreen, 1 mL; Mobile phase A: 0.1 mol/L phosphate buffer +1.8 mol/L sodium sulfate, pH 7.0 ; B: 0.1 mol/L phosphate buffer, pH 7.0; Gradient: 30 min linear; Flow Rate: 1 mL/min; Injection Vol.: 50 μ L; Samples: Ribonuclease A, Lysozyme, γ -Chymotrypsinogen 1 mg/mL

Column: ToyoScreen, 1 mL; Mobile phase A: 0.1 mol/L phosphate buffer +1.8 mol/L sodium sulfate, pH 7.0; B: 0.1 mol/L phosphate buffer, pH 7.0; Gradient: 30 min linear; Flow Rate: 1 mL/min; Injection Vol.: 50 μ L; Samples: Mouse Ascites Fluid: A:B = 1:1:2

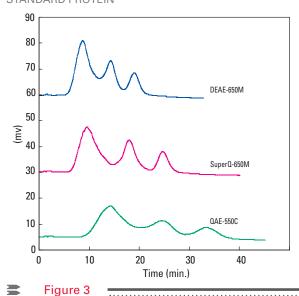
IEX - ION EXCHANGE

Ion exchange chromatography (IEX) separates molecules based on the ionic interaction of the molecule with the charged support. The net surface charge of proteins is dependent on the pH and ionic strength of the mobile phase. The development of optimum chromatographic conditions requires knowledge of both the protein's pI and the pKa of the ion exchange media. In biopurification IEC is used either in 'bind/elute mode' or in 'flow-through mode'. Ion exchange media should be selected according to the properties of the feedstock and the objective of the process step. Factors influencing the final choice are binding capacity of the resin, target scale and speed of the purification step.

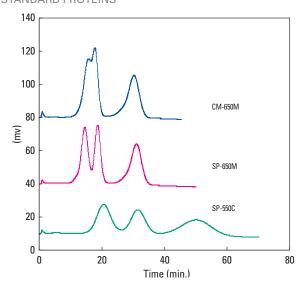
TOYOPEARLIEC resins are available in pore sizes of 1000 Å, 750 Å and 500 Å. TOYOPEARL-550 and -600 resins are designed for the purification of small to medium size proteins. The smaller pores result in increased surface area, thus offering more binding sites and high binding capacities.

ToyoScreen columns are offered in strong and weak functionalities for both cation and anion ligand types. Functional groups comprise of sulfopropyl and carboxymethyl groups for cation exchange or quaternary ammonium or diethylaminoethyl groups for anion exchange resins, respectively.

SREENING OF TOYOPEARL ANION IEC RESINS - STANDARD PROTEIN



SCREENING OF TOYOPEARL CATION IEC RESINS - STANDARD PROTEINS



Column: ToyoScreen, 1 mL; Mobile phase A: 20 mmol/L Tris-HCl, pH 8.0 Mobile phase B: 20 mmol/L Tris-HCl + 0.5 mol/L NaCl, pH 8.0; Gradient: B 0-->100% 60min linear; Flow Rate: 1 mL/min; Samples: Transferrin, Ovalbumin, Trypsin Inhibitor 1 mg/mL each

Column: ToyoScreen, 1 mL; Mobile phase A: 20 mmol/L phosphate buffer, pH 6.0; B: 20 mmol/L phosphate buffer + 0.5 mol/L NaCl, pH 6.0; Gradient: B 0-->100% 60min linear; Flow Rate: 1 mL/min; Samples: α -ChymotrypsinogenA, CytochromeC, Lysozyme 1 mg/mL each

The particle surfaces are modified either by traditional or network bonding chemistries. Network attachment chemistry improves the accessibility of the ligand groups. This significantly improves binding capacity and mass transfer. This technology is applied in TOYOPEARL Giga-Cap and Super Q resins. Figure 3 shows the separation of protein standards for some TOYOPEARL ion exchange chemistries.

MX - MIXED MODE

Mixed mode media combine ion exchange with hydrophobic interaction functionalities. They bind the target based on the hydrophobic interaction and elute the target when ionic interactions more precise electrostatic repulsion takes the lead. The multimodal cation exchanger TOYO-PEARL MX-Trp-650M is salt tolerant and shows unique selectivity towards specific targets.

Since the ionic and hydrophobic properties of the ligand vary with salt concentration and pH, optimization of eluents for adsorption, washing and elution is crucial. Toyo-Screen MX-Trp cartridges are ideally suited to determine proper conditions for this multimodal cation exchanger.

AFC - AFFINITY

In affinity chromatography (AFC), the ligands employed are specific to a particular protein class or functional group on the accessible surface of the target molecule. ToyoScreen affinity columns are offered in four group specific ligand chemistries: AF-rProtein A HC-650F and AF-rProtein A-650F, AF-Blue HC-650M, AF-Chelate-650M, and AF-Red 650M.

AF-rProtein A HC-650F/A-650F is used for the purification of monoclonal antibodies.

AF-Blue HC-650M is specific for kinases, phosphatases, dehydrogenases and other molecules such as albumin and blood coagulation factors.

AF-Red-650M is specific for dehydrogenases and other proteins such as plasminogen.

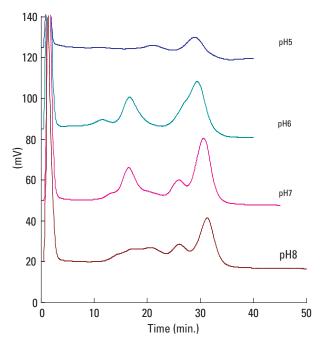
AF-Chelate-650M can be converted to either the Ni⁺⁺, Ca⁺⁺ or Zn⁺⁺ form. When converted to the Ni⁺⁺ form it is an excellent resin for metal ligand affinity for molecules containing His-tags.

ToyoScreen affinity columns allow for the quick assessment of optimum binding conditions for any of these columns.

METHOD OPTIMIZATION

Beyond the determination of selectivity and capacity for the target molecules during resin screening experiments, ToyoScreen columns can be used to quickly establish optimum elution conditions. Varying pH, salt type, salt gradients and flow rate are common experimental parameters explored. The effect of varying salt type and pH are shown in Figure 4 for Anti-TSH in cell culture supernatant on ToyoScreen Phenyl-650M.

EFFECT OF ELUENT PH AND SALT TYPE ON SEPARATION OF CELL CULTURE SUPERNATANT



Column: ToyoScreen Phenyl-650M, 1 mL; Mobile phase A: 0.1 mol/L phosphate buffer +1.8 mol/L ammonium sulfate, pH 7.0; B: 0.1 mol/L phosphate buffer, pH 7.0; Flow Rate: 1 mL/min Gradient: 30 min linear, 30 CV; Injection Vol.: 200 L; Samples: Cell culture supernatant (x4 diluted) (antibody: Anti-TSH)

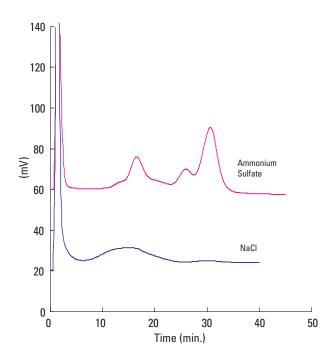


Figure 4

Column: ToyoScreen Phenyl-650M, 1 mL, Mobile phase A: 0.1 mol/L phosphate buffer containing 1.8 mol/L each salt, pH 7.0; B: 0.1 mol/L phosphate buffer, pH7.0; Flow Rate: 1 mL/min Gradient: 30 min linear, 30 CV; Injection Vol.: 200 L; Samples: Cell culture supernatant (x 4 diluted) (antibody: Anti-TSH)

0

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SCALABILITY

Initial results from resin screening and optimization with ToyoScreen columns accurately predict the separation behavior at larger scales. Figure 5, illustrates the similar retention time behavior between 1 mL ToyoScreen columns and conventional 7.5 mm ID x 7.5 cm analytical columns. Additionally, Figure 6 depicts a practical antibody scale up in which conditions were set using a 1 mL ToyoScreen and applied to a 10 mL conventional column with a different inner diameter and length.

COMPARISON OF SELECTIVITY BETWEEN TOYOSCREEN AND CONVENTIONAL COLUMN

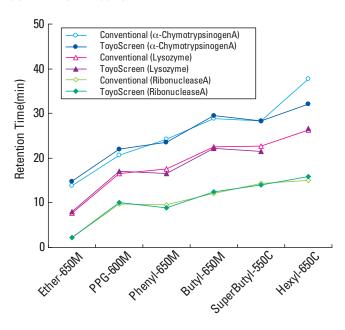
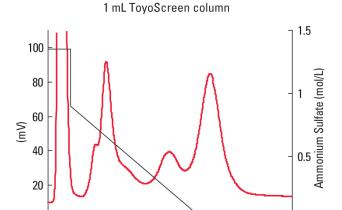


Figure 5

Columns: ToyoScreen (1 mL), Conventional column (7.5 mm ID x 7.5 cm) Mobile phase A: 0.1 mol/L phosphate buffer + 1.8 mol/L sodium sulfate, pH 7.0; B: 0.1 mol/L phosphate buffer, pH 7.0; Gradient: 30 min linear, 30 CV Flow Rate: 1 mL/min; Injection Vol.: 50 μ L; Samples: Ribonuclease A, Lysozyme, α -Chymotrypsinogen; 1 mg/mL

*) Retention time of conventional column was plotted after converting following equation: plotted value = actual measurement value - 4.82

COMPARISON OF CHROMATOGRAMS BETWEEN TOYOSCREEN AND CONVENTIONAL COLUMNS



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Time (min.)

40

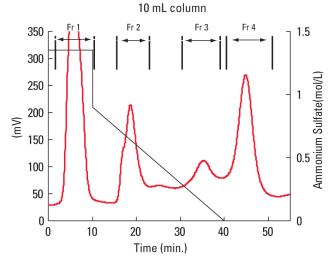


Figure 6

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Packing: TOYOPEARL Phenyl-650M; Mobile phase: (A) 0.1 mol/L phosphate buffer containing 1.8 mol/L (NH $_4$) $_2$ SO $_4$, pH 7.0; (B) 0.1 mol/L phosphate buffer, pH 7.0; Sample: Anti-TSH from cell culture supernatant (x 4 diluted)

Column: 6.4 mm ID x 3 cm L 14.6 mm ID x 6 cm L Injection Volume: 500 μL 5,000 μL Flow Rate: 0.5 mL/min; 0.5 CV/min; 93 cm/hr 2.5 mL/min; 0.25 CV/min; 90 cm/hr Gradient Profile: 25% B; 0 - 5 min (isocratic) 25% B; 0-10 min (isocratic) 50% B: 5 min (step) 50% B: 10 min (step)		1 mL ToyoScreen	10 mL column
Flow Rate: 0.5 mL/min; 2.5 mL/min; 0.5 CV/min; 93 cm/hr 0.25 CV/min; 90 cm/hr Gradient Profile: 25% B; 0 - 5 min (isocratic) 25% B; 0-10 min (isocratic) (isocratic) 50% B: 5 min (step) 50% B: 10 min (step)	Column:	6.4 mm ID x 3 cm L	14.6 mm ID x 6 cm L
0.5 CV/min; 93 cm/hr 0.25 CV/min; 90 cm/hr Gradient Profile: 25% B; 0 - 5 min (isocratic) 25% B; 0-10 min (isocratic) (isocratic) 50% B: 5 min (step) 50% B: 10 min (step)	Injection Volume:	500 μL	5,000 μL
(isocratic) (isocratic) 50% B: 5 min (step) 50% B: 10 min (step)	Flow Rate:	, ,	. ,
	Gradient Profile:	,	,
		50% B: 5 min (step)	50% B: 10 min (step)
50% to 100% B; 50% to 100% B; 5-35 min (linear) 10-40 min (linear)		50% to 100% B; 5-35 min (linear)	50% to 100% B; 10-40 min (linear)
Gradient Slope*: 0.06 mol/L/min 0.012 mol/L/min	Gradient Slope*:	0.06 mol/L/min	0.012 mol/L/min

^{*} The gradient slope is the change in ionic strength per unit volume. Gradient volume is the product of flow rate and gradient time.

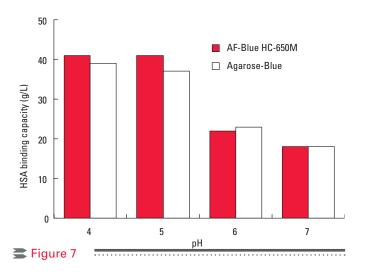
LABORATORY BENCHTOP PURIFICATIONS

Some ToyoScreen affinity columns can be used in simple one step laboratory purifications. This can result in either the isolation of a target molecule or the removal of an overly abundant impurity such as human serum albumin in blood.

REMOVAL OF HUMAN SERUM ALBUMIN

ToyoScreen AF-Blue HC-650M has a very high capacity for HSA as shown in Figure 7. It can be used to remove HSA or to purify albumin conjugated molecules.

COMPARISON OF HUMAN SERUM ALBUMIN BINDING CAPACITIES AT VARIOUS phs OF AF-BLUE HC-650M AND AGAROSE (BLUE FUNCTIONALIZED AGAROSE) RESINS



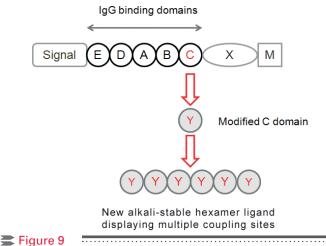
PURIFICATION OF HISTIDINE TAGGED (HIS-TAGGED) PROTEINS

A popular technique for bench scale purification of recombinant proteins is the expression of the protein with a poly-histidine tag and using a chelate column in the Ni⁺⁺ form to selectively bind and elute the fusion protein. The histidine tag is subsequently cleaved from the protein for further work. As shown in Figure 8, ToyoScreen AF-Chelate-650M can be placed into the Ni⁺⁺ form and used to purify his-tagged proteins.

MONOCLONAL ANTIBODY PURIFICATION

ToyoScreen AF-rProtein A-HC 650F has a new recombinant protein A ligand attached to it as shown in Figure 9. It has an extremely high binding capacity. It is very base stable and can be cleaned with 0.1 - 0.5 mol/L NaOH. Figure 10 shows the high IgG binding capacities measured at various linear velocities and various feedstock concentrations. The high capacity at short residence times enables efficient small scale purifications for R&D purposes with ToyoScreen cartridges.

TOYOPEARL AF-rPROTEIN A HC-650F LIGAND STRUCTURE



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PURIFICATION OF HIS-TAG NusG FUSION PROTEIN

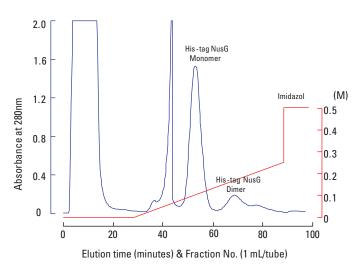
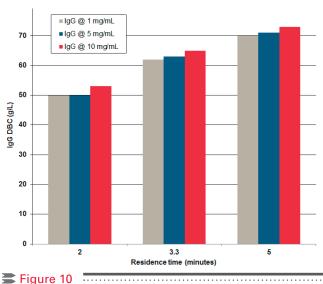


Figure 8

Column: ToyoScreen Chelate-650M (Ni-chelate), 5 mL; Starting Buffer: 20 mmol/L NaPi, pH 8.2, 0.01%NaN $_3$; Buffer B: 0.5 mol/L Imidazol, 20 mmol/L NaPi, pH 7.4, 0.01% NaN $_3$; Flow rate: 1 mL/min, at RT, Gradient: 0.25 min= 0% B, 25.85 min=0.50% B, 85.90 min=100%, 90.100 min=0% B; Sample: crude cell extract (5 mL, 10 mg, 20 mmol/L NaPi, pH 8.2, 0.01%NaN $_3$)

DBC AT VARIOUS LOADS AND RESIDENCE TIMES



Column: TOYOPEARL AF-rProtein A HC-650F (5 mm ID \times 5 cm); Mobile phase: 20 mmol/L sodium phosphate, 150 mmol/L NaCl pH 7.4; Residence time: 2, 3.3, 5 min; Detection: UV @ 280 nm; Sample: polyclonal human IgG @ 1, 5, 10 g/L in mobile phase; DBC measured at 10 % breakthrough

Ordering information

ToyoScreen®

Part-No Description	Comment	Part-No Description Comment
ION EXCHANGE		AFFINITY
0021360 ToyoScreen DEAE-650M	1 mL x 6 each	0023430 ToyoScreen AF-rProtein A HC-650F 1 mL x 5 each
0021361 ToyoScreen DEAE-650M	5 mL x 6 each	0023430 ToyoScreen AF-Protein A HC-650F 5 mL x 1 each
0021362 ToyoScreen SuperQ-650M	1 mL x 6 each	0023431 ToyoScreen AF-rProtein A HC-650F 5 mL x 5 each
0021363 ToyoScreen SuperQ-650M	5 mL x 6 each	0022809 ToyoScreen AF-rProtein A-650F 1 mL x 5 each
0021364 ToyoScreen QAE-550C	1 mL x 6 each	0022810 ToyoScreen AF-rProtein A-650F 5 mL x 1 each
0021365 ToyoScreen QAE-550C	5 mL x 6 each	•
,	1 mL x 6 each	
0021859 ToyoScreen GigaCap Q-650M		0021384 ToyoScreen AF-Chelate-650M 1 mL x 6 each
0021860 ToyoScreen GigaCap Q-650M	5 mL x 6 each	0021385 ToyoScreen AF-Chelate-650M 5 mL x 6 each
0021366 ToyoScreen CM-650M	1 mL x 6 each	0021386 ToyoScreen AF-Blue HC-650M 1 mL x 6 each
0021367 ToyoScreen CM-650M	5 mL x 6 each	0021387 ToyoScreen AF-Blue HC-650M 5 mL x 6 each
0021368 ToyoScreen SP-650ML	1 mL x 6 each	0021388 ToyoScreen AF-Red-650M 1 mL x 6 each
0021369 ToyoScreen SP-650M	5 mL x 6 each	0021389 ToyoScreen AF-Red-650M 5 mL x 6 each
0021370 ToyoScreen SP-550C	1 mL x 6 each	0021390 ToyoScreen AF-HeparinHC-650M 1 mL x 6 each
0021371 ToyoScreen SP-550C	5 mL x 6 each	0021391 ToyoScreen AF-HeparinHC-650M 5 mL x 6 each
0021868 ToyoScreen GigaCap S-650M	1 mL x 6 each	
0021869 ToyoScreen GigaCap S-650M	5 mL x 6 each	MIX PACKS
0021870 ToyoScreen MegaCap II SP-550EC	1 mL x 6 each	0021392 ToyoScreen IEC Anion Mix Pack 1 mL x 5 Grades
0021871 ToyoScreen MegaCap II SP-550EC	5 mL x 6 each	(DEAE-650M, SuperQ-650M, QAE-550C, x 1 each
0021992 ToyoScreen Q-600C AR	1 mL x 6 each	GigaCap Q-650M, Q-600C AR)
0021923 ToyoScreen Q-600C AR	5 mL x 6 each	0021393 ToyoScreen IEC Anion Mix Pack 5 mL x 5 Grades
0023443 ToyoScreen NH ₂ -750F	1 mL x 6 each	(DEAE-650M, SuperQ-650M, QAE-550C, x 1 each
0023444 ToyoScreen NH2-750F	5 mL x 6 each	GigaCap Q-650M, Q-600C AR)
		0021394 ToyoScreen IEC Cation Mix Pack 1 mL x 5 Grades
HYDROPHOBIC INTERACTION		(CM-650M, SP-650M, SP-550C, , x 1 each
0021372 ToyoScreen Ether-650M	1 mL x 6 each	GigaCap CM-650M, GigaCap S-650M)
0021373 ToyoScreen Ether-650M	5 mL x 6 each	0021395 ToyoScreen IEC Cation Mix Pack 5 mL x 5 Grades
0021374 ToyoScreen Phenyl-650M	1 mL x 6 each	(CM-650M, SP-650M, SP-550C, GigaCap CM-650M, x 1 each
0021375 ToyoScreen Phenyl-650M	5 mL x 6 each	GigaCap S-650M)
0021376 ToyoScreen Butyl-650M	1 mL x 6 each	0021396 ToyoScreen IEC Mix Pack 1 mL x 6 Grades
0021377 ToyoScreen Butyl-650M	5 mL x 6 each	(GigaCap Q-650M, SuperQ-650M, Q-600 AR, x 1 each
0021378 ToyoScreen Hexyl-650C	1 mL x 6 each	GigaCap CM-650M, GigaCap S-650M, SP-550C)
0021379 ToyoScreen Hexyl-650C	5 mL x 6 each	0021397 ToyoScreen IEC Mix Pack 5 mL x 6 Grades
0021380 ToyoScreen PPG-600M	1 mL x 6 each	(GigaCap Q-650M, SuperQ-650M, Q-600 AR, x 1 each
0021381 ToyoScreen PPG-600M	5 mL x 6 each	GigaCap CM-650M, GigaCap S-650M, SP-550C)
0021892 ToyoScreen Phenyl-600M	1 mL x 6 each	0021398 ToyoScreen HIC Mix Pack 1 mL x 6 Grades
0021893 ToyoScreen Phenyl-600M	5 mL x 6 each	(PPG-600M, Phenyl-600M, Phenyl-650M Butyl-600M, x 1 each
0021494 ToyoScreen Butyl-600M	1 mL x 6 each	Butyl-650M, Hexyl-650C)
0021495 ToyoScreen Butyl-600M	5 mL x 6 each	0021399 ToyoScreen HIC Mix Pack 5 mL x 6 Grades
0021382 ToyoScreen SuperButyl-550C	1 mL x 6 each	(PPG-600M, Phenyl-600M, Phenyl-650M, x 1 each
0021383 ToyoScreen SuperButyl-550C	5 mL x 6 each	Butyl-600M, Butyl-650M, Hexyl-650C)
MIXED MODE		TOYOSCREEN COLUMN ACCESSORIES
0022824 ToyoScreen MX-Trp-650M	1 mL x 6 each	0021400 ToyoScreen Column Holder
0022825 ToyoScreen MX-Trp-650M	5 mL x 6 each	
		Please note that all of the Toy Screen packages shown above
		require a ToyoScreen column older PN 21400 for operation.
		and a second