

Application Note



PHOTODEGRADABLE POLYMER DEGRADATION ANALYSIS USING THE EcoSEC[®] GPC SYSTEM

INTRODUCTION

Photoresponsive polymers have several advantages over other stimuli responsive materials due to the spatial and temporal control of the input.¹ Photoactivation can be used to influence various polymer properties such as release or capture of additives, change in viscosity, modulus and pH. Early efforts in the field of photoresponsive polymers were aimed primarily at decreasing the environmental impact of plastics in landfills and on marine life. However, the long degradation time and non-biodegradability of these materials make them unsuitable for biological applications. Currently there is a strong need in biomedical applications for polymers that are both photodegradable and biodegradable. Such polymers are being investigated as drug delivery devices and as platforms with phototunable physical and mechanical properties.

To address the needs of biological application, a new class of photodegradable polycarbonate materials has been developed based on the alkoxyphenacyl photoactive moiety that undergo controlled degradation to oligomers upon irradiation at 300 nm.¹ These polycarbonates are mechanically robust, biodegradable, and stable at high temperatures in the absence of light. The combination of the thermal and mechanical properties of these polymers promises usefulness in biomedical applications such as controlled drug release devices, ocular implants, and dermal patches.

The photodegradable polycarbonate homopolymer as well as different copolymers with poly(ethylene glycol) (PEG) were synthesized and characterized by gel permeation chromatography (GPC) using the EcoSEC GPC System. Here we report, through molar mass averages and polydispersity index values as determined by GPC, the photodegradation of the polycarbonate homopolymer, 5% PEG copolymer, and 10% PEG copolymer by irradiation of the polymers in chloroform in a Rayonet reactor at 300 nm as well as the hydrolytic degradation of the copolymers by incubation in phosphate buffered saline (PBS) at 37°C.

EXPERIMENTAL CONDITIONS

Analysis of the homopolymer, copolymers, photodegraded samples, and hydrolytically degraded samples were performed on an EcoSEC GPC System equipped with a dual flow refractive index detector (RI) and UV detector. The polymers under investigation have UV absorption from 250 to 320 nm with a λ_{max} at 280 nm. Separation occurred over a column bank consisting of two 6 mm ID × 15 cm, 3 µm TSKgel[®] SuperH3000 (exclusion limit 6 × 10⁴ g/mol, PN 0017993) and one 6 mm ID × 15 cm, 3 µm TSKgel SuperH4000 (exclusion limit 5 × 10⁵ g/mol, PN 0017994) columns. The mobile phase and solvent were chloroform (CHCl₃) at flow rate of 0.38 mL/min. Detector, pump oven, and column oven were maintained at 40°C. For all chromatographic determinations, results are those based on a polystyrene calibration curve.

POLYMER PHOTODEGRADATION WITH TIME OF IRRADIATION

The photodegradable homopolymer poly(2-hydroxy-1-(4-(3-hydroxypropoxy)phenyl)ethanonecarbonate) (20 mg) was dissolved in chloroform and transferred to a quartz tube. The sample was irradiated in a Rayonet RPR-200 reactor at 300 nm where the polymer has a significant UV absorption (5.34 mW/cm²). Every 5 minutes, the reactor was turned off and 1 mL of reaction mixture was taken out and filtered through a 0.45 μ m PTFE filter. A total of 9 samples were taken in 40 minutes of irradiation and the rest of the solution was irradiated for an extra hour.

HYDROLYTIC DEGRADATION OF POLYMERS

Poly(2-hydroxy-1-(4-(3-hydroxypropoxy)phenyl)ethanone carbonate)-co-poly((poly(ethyleneglycol)diol)carbonate) copolymers were dissolved in CHCl₃. The solution was stirred for a few hours to allow the polymer to dissolve completely. Polymer films were prepared by solvent casting the above solution into a Teflon[®] dish and allowing the solvent to evaporate slowly overnight and were dried under vacuum prior to use. Hydrolytic degradation was monitored by immersing half of each polymer film (~36 mg) in a vial containing 5 mL of PBS solution in an incubator at 37°C. At the end of every week (total of 4 weeks) a small piece of film was taken from the vial, dried, dissolved in CHCl₃ and analyzed by GPC.

SYNTHESIS OF PHOTODEGRADABLE POLYMERS¹

Figure 1



(i) K₂CO₂/18-crown-6, acetone reflux. (ii) CuBr₂, EtOH, CHCl₃ (over two steps: 75% yield).
(iii) NaOAc, CH₃COOH, H₂O (99% yield). (iv) 1. NaOH, MeOH, 2. NaHSO₄ (46% yield).
(v) Triphosgene, CHCl₃, pyridine, PEG_{1k}

MOLAR MASS DISTRIBUTIONS AND POLYDISPERSITY INDEX

Composition	Mn (g/mol)	Mw (g/mol)	PDI
Homopolymer	1.29 x 10 ⁴	2.95 x 10 ⁴	2.3
5% PEG	2.27 x 10 ⁴	2.63 x 10 ⁴	1.2
10% PEG	8,810	1.04 x 10 ⁴	1.2
Table 1			

RESULTS AND DISCUSSION

Polycarbonate homopolymer and copolymers were synthesized using the scheme shown in Figure 1. The polystyrene relative molar mass averages, Mn and Mw, and the polydispersity index, PDI, were determined using the EcoSEC GPC System with dual flow RI and UV detectors. Three polymers were synthesized and analyzed: an alkoxyphenacyl-based polycarbonate homopolymer, 5% PEG copolymer, and 10% PEG copolymer. The polystyrene relative molar mass averages, Mn and Mw, and the polydispersity index, PDI, for the initial homopolymer and copolymers are given in Table 1. The polystyrene weight-average molar mass values were also determined for the homopolymer and copolymers after photochemical and hydrolytic degradation.

The photochemical degradation of the homopolymer and copolymers was examined by irradiation in $CHCl_3$ in a Rayonet reactor at 300 nm. GPC elution profiles of the irradiated homopolymer showed that the polymer underwent controlled time-dependent chain scissions upon irradiation, Figure 2. Prior to irradiation the polystyrene relative weight-average, molar mass was determined to be 2.95 × 10^4 g/mol. Within five minutes of irradiation, there was a loss of three-fourths of the Mw,- of the polymer, Mw= 7.2 × 10^3 g/mol. After a hundred minutes of irradiation of the polymer the weight-average molar mass, Mw, of the polymer had decreased to 146 g/mol. The Mw values were shown to continuously decrease with irradiation time. The GPC elution profile shifts towards longer retention times with increasing irradiation time, indicating that the irradi-

DECREASE IN Mw-WITH IRRADIATION TIME FOR THE POLYCAR-BONATE HOMOPOLYMER



ated homopolymer is smaller in polymer size compared to the non-irradiated homopolymer. The photodegradation of the copolymer polymers showed a similar trend to that of the homopolymer.

Hydrolytic degradation of the polymers was obtained by incubation of the copolymers, 5% and 10% PEG, in phosphate buffered saline at 37°C. For the copolymers, hydrolytic degradation is reflected by the molar mass loss with the time of incubation, Figure 3. For example, over a period of 28 days, the copolymer with 5% PEG showed increasing molar mass loss with time and ultimate loss of 61% of molar mass on day 28. As expected, the photochemical degradation is much faster – the polymer undergoes almost complete photodegradation within 30 minutes, Figure 2, while hydrolytic degradation over 28 days results in a 61% loss for the 5% PEG copolymer, Figure 3.





CONCLUSIONS

The synthesis and properties of a new class of photodegradable polymers that undergo controlled chain scission upon irradiation at 300 nm were analyzed using the EcoSEC GPC System. The polystyrene relative molar mass averages of a homopolymer and two copolymers were determined before, during and after photochemical and hydrolytic degradation. Additionally, the GPC elution profile was monitored during both photodegradation processes. The photodegradation results demonstrate that the polymers quickly lose their molar mass upon irradiation. These properties along with others determined via different characterization methods, e.g. NMR, make these polycarbonate homopolymer and copolymers valuable for many biomedical applications.¹ The low dispersion design of the EcoSEC GPC System combined with the use of 15 cm long TSKgel columns provided a fast and robust method for monitoring the degradation by GPC.

REFERENCES

1. Sun, S.; Chamsaz, E.A.; Joy A. Macro Lett., 2012 1 (10), 1184-1188